# **Indolo[3,2-b]quinolines: Synthesis, Biological Evaluation and Structure Activity-Relationships**

Eyunni V.K. Suresh Kumar, Jagan R. Etukala and Seth Y. Ablordeppey\*

*Division of Basic Pharmaceutical Sciences, College of Pharmacy & Pharmaceutical Sciences, Florida A&M University, Tallahassee, FL. 32309, USA* 

**Abstract:** The tetracyclic indolo[3,2-b]quinoline ring system constitutes an important structural moiety in natural products exhibiting numerous biological activities. In particular, indolo [3, 2-b]quinoline, commonly known as linear quindoline is of particular interest, because of its rigid structure and scope of derivatization. Although the core linear quindoline skeleton shows little or no activity in several biological systems, introduction of a methyl group on the N-5 atom leading to cryptolepine induces remarkable activity against a broad spectrum of biological targets. A number of analogs of quindoline and cryptolepine have been synthesized, incorporating various functional groups on the core quindoline skeleton leading to improved biological activities. In this review, we describe various synthetic methodologies leading to the quindoline scaffold, the biological activities and the structure activity relationships (SAR) of quindoline derivatives toward different disease states to give a better picture of the importance of this moiety in medicinal chemistry.

**Key words:** Cryptolepine, indoloquinolines, synthesis, structure-activity relationship (SAR) studies.

# **INTRODUCTION**

 Cryptolepine or 5-methyl-5H-indolo[3, 2-b] quinoline (**1**) is an example of a natural product whose synthesis was reported prior to its isolation from plants. The first synthesis was reported in 1906 by Fichter and coworkers [1] before its isolation from *Cryptolepis triangularis* N. E. Br by Clinquart in 1929 [2] and again by Delvaux [3] soon thereafter. Cryptolepine has also been isolated as the major alkaloid from Sida sp. (Malvaceae) medicinal plants used by indigenous practitioners of Sri Lanka [4].

 However, cryptolepine was first isolated from the shrub *Cryptolepis sanguinolenta* by Gellert from root samples obtained in Nigeria [5] and again by Dwuma-Badu and coworkers from root samples obtained from Ghana [6]. In traditional folk medicine of West and Central Africa, a decoction of the root of the plant *C. sanguinolenta* is used to treat fevers (including fever caused by malaria), upper respiratory infections and venereal diseases. Apart from cryptolepine, neocryptolepine {cryptotackieine, 5-methyl-5H-indolo[2, 3 b]quinoline} (**2**), isocryptolepine {cryptosanguinolentine, 5 methyl-5H-indolo[3, 2-c]quinoline} (**3**), quindoline (**4**) and several other isomeric indoloquinolines complete the 13 characterized alkaloids isolated from this plant Fig. (**1**).

 Under acidic conditions, these tetracyclic compounds exist in their salt forms possessing a broad spectrum of biological activities. However, under basic conditions, the N-5 atom in these alkaloids assumes  $sp<sup>3</sup>$  hybridized form and the alkaloids exist as free bases as represented (**5**). Thus, a quaternary nitrogen is able to undergo a reversible acid – base transformation from yellow to pink respectively Fig. (**2**).

 Biological testing of Cryptolepine has led to its identification as having a broad spectrum of activities including antibacterial, [7] antimalarial, [8] antifungal, [9] antithrombotic, [10] vasodilation, [11] antihyperglycemic, [12] anti cancer and cytotoxic, [13] anti-inflammatory, [14] hypotensive and antipyretic, [15] presynaptic  $\alpha$ -adrenoceptor blocking action, [16] and anti-muscarinic [17] properties. The mechanism by which cryptolepine produces these biological activities is not completely known. However, several lines of evidence suggest that DNA intercalation and stabilization of topoisomerase II-DNA complex may play some role [18]. As a result of these interesting biological activities, many groups world-wide have designed new methodologies to synthesize derivatives of quindoline (**4**) and have attempted to study the structure activity relationship of these indoloquinolines towards various disease states. Thus, the purpose of this review was to assemble the various synthetic strategies leading to the construction of the indoloquinoline structure as well as the biological activities and structure-activity relationships of the indoloquinoline alkaloids to date.

# **METHODS OF CHEMICAL SYNTHESES OF INDOLO[3,2-B]QUINOLINE** (**QUINDOLINE**)

 Several routes are now available for the synthesis of cryptolepine analogues. The most versatile method reported by Holt and Petrow [19] was based on the synthesis of linear quindoline (**4**) in which isatin (**6**) is condensed with 3 acetylindoxyl (**7**) under basic and oxygen-free conditions and the resulting quindoline-11-carboxylic acid (**8**) is decarboxylated by heating in diphenylether or mineral oil to yield **4**. (Scheme **1**).

 Better yields are obtained by using indoxyl-1, 3-diacetate (**9**) Fig. (**3**) as starting material instead of **7**. However, strongly electron withdrawing substituents on isatin such as a nitro group inhibit the first step in the synthesis.

 Quindoline can be methylated at the N-5 position by methyl iodide using the method described by Fichter [1] to

**<sup>\*</sup>**Address correspondence to this author at the Division of Basic Pharmaceutical Sciences, College of Pharmacy & Pharmaceutical Sciences, Florida A&M University, Tallahassee, FL. 32309, USA; Tel: 850-599-3834; Fax: 850-599-3934E-mail: seth.ablordeppey@famu.edu



**Fig. (1).** Indoloquinolines.



**Fig. (2).** Reversible transformation of Cryptolepine under acidic and basic conditions.

yield cryptolepine as the hydroiodide salt (Scheme **2**). Another method, developed by Bierer and coworkers [20] for large scale reactions involves the use of methyl triflate as the alkylating agent to obtain hydrotriflate salts of cryptolepine. Conversion from the hydrotriflate or hydroiodide salts to other salt forms can be achieved by reaction with a base and treatment of the resulting free base with ethereal HCl solution for example, to provide cryptolepine hydrochloride salt. Hence, the quaternary N-5 atom does not behave in the normal way as a permanent positively charged atom. These acid-base changes are also associated with color changes. While cryptolepine free base (**5**) is deep purple, the salt form (**1a**) is yellow or orange depending on the solvent and pH of the solution (Scheme **2**).

 A major disadvantage of the method developed by Holt and others is the long reaction times  $( \geq 10 \text{ days})$  during the first step of the reaction, i.e., preparation of the quindoline-11-carboxylic acid (**8**) (Scheme **1**). To reduce the reaction times and generate quindoline rapidly, we developed a new and more efficient synthesis of quindoline [21] which requires only two steps and was completed in one day with an overall yield of 22 %. The synthesis involved N-arylation of 3-aminoquinoline (**10**) with triphenyl-bismuth diacetate in

the presence of metallic copper to yield 3-anilinoquinoline (**11**) in excellent yield. A similar procedure using triphenylbismuth in the presence of copper diacetate also produced **11** but with low yields. Cyclization of **11** was effected by palladium (II) acetate in refluxing trifluoroacetic acid to afford **4** (Scheme **3**). The cyclization reaction was however low yielding because of the formation of angular quindoline in addition to linear quindoline. The ratio of the regioisomers however, depends on the substituents on the rings (Personal observations). Separation of the regioisomers is easily achieved by flash chromatography.



**Fig. (3).** Structure of Indoxyl-1, 3-diacetate.

 More recently, a similar method was reported albeit in low yields, wherein aniline was condensed with 3-bromoquinoline (**12**) resulting in intermediate (**13**) which on oxida-



Reagents: i. KOH,  $N_2$ , 10 days; ii. Mineral Oil/Ph<sub>2</sub>O, 250-300 °C, 2 hrs.

**Scheme 1.** 

**540** *Mini-Reviews in Medicinal Chemistry,* **2008***, Vol. 8, No. 6 Kumar et al.* 



## **Scheme 3.**

tive photocyclization in the presence of traces of iodine, afforded angularly-fused quindoline (**14**) as the major product (51 %) and **4** as the minor product (16 %) [22] (Scheme **4**).

-nucleophilic substitution of 1-phenylsulfonyl-2-acylindole (Scheme **5**).

 Cooper [23] reported the synthesis of hydroxycryptolepine, cryptolepine and quindoline utilizing the intramolecular

 In this process, 1-phenylsulfonylindole (**15**) was lithiated at the 2 position, reacted with 2-nitrobenzaldehyde (**16**) and the resulting alcohol was oxidized using MnO<sub>2</sub>. Subsequent



**Scheme 4.** 





**Scheme 5.** 

catalytic reduction of the nitro group and N-benzoylation gave amido-ketone (**17**). N-Deprotonation using NaH allowed ring closure to **18** and hydrolysis produced **19**. Heating **19** in POCl3 produced **20** and catalytic hydrogenolysis gave quindoline (Scheme **5**).

 Recently, Ho [24].reported a synthetic procedure leading to the indolo[3,2-b]quinoline system and the isomeric indolo[2,3-b]quinoline scaffold (**21**) Fig. (**4**), based on consideration of their accessibility from a common symmetrical intermediate, 1, 3-bis(1-nitrophenyl)propan-2-one (**22**) which was readily obtained from (2-nitrophenyl)acetic acid (**23**).



**Fig. (4).** Structure of Indolo[2, 3-b]quinoline.

 Reduction of compound **22** with sodium dithionite in aqueous methanol led to the diamino ketone which underwent spontaneous cyclization to produce the first cyclized product **24**. Transformation of **24** to **4** was achieved with iodosobenzene diacetate as the oxidizing agent (Scheme **6**).

 In another synthetic procedure reported by Radl and coworkers [25], 2-nitro phenacyl bromide (**25**) was reacted with ethyl-2-cyanophenylcarbamate (**26**) under basic conditions to yield compound **27**. Using the nitro group as a leaving group, cyclization of **27** under basic conditions yielded quindolinone (**19**) which can be converted to quindoline, as shown in Scheme **7**.

 More recently, Dutta and coworkers [26] reported a simple method for the synthesis of various 2-substituted quindolines starting from 2-nitroacetophenone (**28**) as shown in Scheme **8**. The important steps involved in the synthesis are regioselective thermal cyclization of enaminoimine hydrochloride and generation of a nitrene intermediate followed by its insertion into a  $sp<sup>2</sup>$  C-H bond. The first step in this process involves treatment of 2-nitroacetophenone (**28**) with POCl<sub>3</sub> in DMF and a Vilsmeier-Haack reaction to produce  $\beta$ chlorocinnamaldehyde, (**29**) with the nitro group remaining intact. At 0 °C, chloroaldehyde (29) reacts with aniline in ethanolic HCl to produce the corresponding enaminoimine hydrochloride (**30**) in a very good yield. Thermal cyclization of 30 at 200-250 °C, produced 2-(2-nitrophenyl)quinoline (**31**) as the major isolable product. The desired quindoline was synthesized by heating 31 with triethyl phosphate at 160 <sup>o</sup>C. This step involves intramolecular annulation through a nitrene intermediate.

 Bierer [27] synthesized a methoxy analog of the indoloquinoline by condensation of 1-acetyl-3-oxoindole (**32**) with 2-nitro-3-methoxybenzaldehyde (**33**) using a catalytic amount of piperidine to give compound **34** as a mixture of E/Z iso-



Reagents: i. DCC, THF, DMAP,  $\Delta$ ; ii. sodium dithionate, MeOH D; iii. PhI(OAc)<sub>2</sub>, THF, RT.

**Scheme 6.** 



Reagents: i. K<sub>2</sub>CO<sub>3</sub>/NaH, DMF, RT; ii. NaH, THF, RT; iii. PCl<sub>5</sub>,  $\Delta$ ; iv. EtOH.HCl, 10 % Pd-C, MeOH

**Scheme 7.** 



Reagents: i. POCl<sub>3</sub>, DMF, 0 °C, 1 h then 80 °C, 4h; ii. 2 N ethanolic HCl, arylamine, 0 °C; iii. heat, 200-250 °C, 5 min; iv. P(OEt)<sub>3</sub>,  $\Delta$ , 4 h;

### **Scheme 8.**

mers. Hydrogenation of **34** gave quindoline, **35** in 78 -85% yields after chromatography. Hydrolytic cleavage of the acetate group in **35** using KOH, yielded the substituted quindoline (**36**) in good yield (Scheme **9**). Unsubstituted quindoline can also be prepared in principle using the same procedure.

 Arzel and co-workers [28] reported a short synthetic route leading to quindoline involving the first reported halogen-dance reaction in the indoloquinoline series. Later, they extended that methodology to synthesize various derivatives of quindoline and cryptolepine towards their studies on antiplasmodial and antitrypanosomal activities of cryptolepine [29] (Scheme **10**). Thus, palladium-catalysed cross-coupling reaction (Suzuki coupling) between iodoquinoline (**37**) and boronic acid (**38**) afforded the biaryl compound (**39**) in high yields. Cyclization to quindoline was achieved by heating **39** in boiling pyridinium chloride and hydrolyzing the resulting N-protected quindoline.

 A widely used method utilizing anthranilic acid (**40**) as starting material was reported by Bierer in 1998 [27]. The approach involved treating anthranilic acid with bromoacetyl bromide to afford the condensation product (**41**) which was then reacted with aniline to give compound **42**. Acid promoted cyclization of **42** with polyphosphoric acid led to compound **19** which on treatment with phosphorous oxy-



Reagents: i. Piperidine (cat), Toluene/CHCl<sub>3</sub>, 4 A<sup>o</sup>, MS. ii. H<sub>2</sub>, Pd/C, MeOH, iii. KOH

**Scheme 9.** 





**Scheme 10.** 



Reagents: i. Bromoacetyl bromide, DMF/dioxane, ii. Anilline, iii. PPA, iv. POCl<sub>3</sub>, v. H<sub>2</sub>, Pd/C, MeOH

#### **Scheme 11.**

chloride gave 11-chloro quindoline (**20**). Hydrogenation of the same on Pd/C led to quindoline in good yields (Scheme **11**).

 Using this approach to synthesize halogenated derivatives however, can be problematic. For example, when the 11-chloro quindoline (**20**) has additional halogen on any other position in the ring, hydrogenation can lead to the stripping of all the halogens on the ring system [27, 30-33]. Thus, although this procedure is widespread and useful for synthesizing substituted quindolines, it is not as useful as some of the other methods when used for halogenated analogs.

# **BIOLOGICAL ACTIVITIES OF INDOLOQUINO-LINES**

 As noted in the introduction, cryptolepine is by far the most popular and most researched indoloquinoline. In the following sections, several of the biological activities associated with the indoloquinoline scaffold are reviewed.

#### **A**. **Antibacterial Activity**

 Bacterial infections such as those associated with *Mycobacterium tuberculosis* and methicillin- and vancomycinresistant *Staphylococcus aureus* (MRSA & VRSA) are of major concern especially for immuno-compromised patients. Given the vast numbers of bacteria, their short generation times and typical gene mutation frequencies of 1 in  $10^7$  bacteria, resistance is inevitable. The mechanism of resistance development in these bacteria may include any one or combination of a) destruction of antibiotic by bacterial enzymes, b) bacterial reprogramming of the antibiotic to lowered susceptibility and c) pumping out antibiotics *via* transmembrane efflux pumps and keeping antibiotic concentration within the bacterial cell below toxic threshold concentrations [34]. To solve the problem of resistance development associated with the treatment of bacterial infections there are three broad strategies. The first and foremost is to develop new antimicrobial agents with a broad spectrum of activity including the production of a super drug that has more than one target in the offending pathogen. Secondly, the search for novel screening approaches and therapeutic agents that have different mechanisms of action from the antibacterials currently

available. Thirdly, containment of resistance by avoiding its emergence and preventing its transmission [35]. Certainly, there is a continuing need for a pipeline of new agents to combat multi-drug resistant (MDR) and new emerging opportunistic bacteria.

 Bioactive plant extracts or constituents are rich sources for development of chemotherapeutic agents in the management and treatment of opportunistic and resistant antibacterial infections. Many plant-based antimicrobials have produced remarkable antibacterial activity [36] and plant sources of indoloquinoline alkaloids are notable in this regard. In 1979, Boakye-Yiadom and others [37] reported antibacterial activity of cryptolepine. It was noted that this activity was both bacteriostatic and bactericidal against *Staphylococcus aureus (S. aureus).* They further speculated that cryptolepine may act in this manner, by causing lysis to *S. aureus*. It was also reported that cryptolepine was more active against Gram-positive bacteria (MIC  $\leq$  100  $\mu$ g/mL) than Gram-negative ones [38] and its killing effect on *S*. *aureu*s was subsequently studied [39]. When cryptolepine was exposed to the bacteria, lysis was observed and the surface morphological appearance of the cells was also altered. A similar effect on morphology and survival of other bacteria and fungi *viz*., *Escherichia coli*, *Candida albicans* and *Saccharomyces Cerevisiae* was observed [40, 41]. Both the hydrochloride salt of cryptolepine and its free base form were assessed against M. *fortuitum* (an alternative screening model to *M. tuberculosis*) and the salt form was found to be ten-fold less active than the free base [38b]. Recent screening of cryptolepine hydrochloride salt against a panel of six species of fast growing mycobacteria showed a significant activity against M. *aurum* and M. *Phlei* (MIC = 2 and 4 g/mL) as compared to the standard antibiotics Ethambutol (1 g/mL) and Isoniazid (1g/mL). Activity against M*. fortuitum* was shown to be moderate (MIC = 16  $\mu$ g/mL) as compared to Ethambutol (8  $\mu$ g/mL) and Isoniazid (0.5  $\mu$ g/ mL) [42]. Another recent report indicates that novel analogs of cryptolepine (**43-46,** Fig. (**5**)) displayed rather broad antimicrobial spectra [43]. Compounds **43**, **45** and **46** showed more expansive antimicrobial/antiparasitic spectra than cryptolepine. In particular, activity against methicillin-resistant *Staphylococcus aureus* (MRSA) was significant. The  $IC_{50}$ values were reported to be 2.0, 3.0 and 6.0  $\mu$ g/mL respec-



**Fig. (5).** New Cryptolepine analogs with antibacterial activity.

tively for compounds  $43$ ,  $45$  and  $46$  against the standard  $IC_{50}$ value of  $0.15 \mu g/mL$  for ciprofloxacin. All four compounds showed significant potency against *Mycobacterium intracellulare*. However, only **45** showed significant potency against *Pseudomonas aeruginosa* ( $IC_{50} = 1.5 \mu g/mL$ ) as compared to an IC<sub>50</sub> value of 0.06  $\mu$ g/mL for ciprofloxacin [43].

### **B**. **Antiprotozoal Activity**

 Malaria continues to be a major cause of morbidity and mortality in developing nations. A primary factor for this is related to the increasing prevalence of *P. falciparum* resistant to chloroquine and other antimalarial agents. In one study, it was found that an 11-fold increase in mortality in children under the age of 4 years occurred within 6 years of the emergence of chloroquine-resistance [44]. Another insect borne disease, African sleeping sickness, also known as trypanosomiasis, is an infection endemic to Sub-Saharan Africa. Although drugs like Pentamidine and Suramin are used in initial stages of treatment and Melarsoprol was used in the second phase of the disease, no agent has been found to treat the later phases of the disease. These problems have spurred a considerable interest in identifying antiprotozoal agents from plant sources which can be delivered orally. Thus, many plants, especially those reputed as traditional medicines have been evaluated [45].

 Wright [8a] has recently published a review article that discussed the potential of cryptolepine analogs as antimalarial drugs. Cryptolepine and its hydrochloride salt (**1b**) were found to have potent *in vitro* activity against various *P. falciparum* strains [8c, 8e, 8f, 46, 47]. Cimanga and coworkers [8f] studied the antiplasmodial activity of cryptolepine, its hydrochloride, 11-hydroxycryptolepine (**47**) and quindoline, and found that both cryptolepine and its hydrochloride salt showed strong *in vitro* antiplasmodial activity against *P. falciparum* chloroquine-resistant strains, K-1 and W-2. However, quindoline was much less active. *In vivo* tests in infected mice showed cryptolepine hydrochloride as having significant activity against *P. berghei yoelii* and *P. berghei berghei*. 11-Hydroxycryptolepine and quindoline were devoid of activity in these assays.

 The influence of the structural modifications at N-5 and C-11 positions of the indoloquinoline moiety against *T. cruzi* Tulahuen strain and chloroquine-resistant K-1 strain of *P*. *falciparum* were studied [29]. A methyl substituent at the N-5 position in the ring seems to be essential for activity since quindoline was about 100 times less active against both para-

sites than cryptolepine. The quindoline dimer **48** showed better activity than quindoline but its index of selectivity (IS) remained in a similar range. Introduction of alkyl groups at the C-11 position alone on the quindoline ring had no significant influence on activity, except that 11-ethyl quindoline (**49**) was about 10 times more active than quindoline against *T. cruzi* trypomastigote forms. With the 5-methyl substituent intact, alkyl groups at the C-11 position reduced activity against *P. falciparum*, and intracellular multiplication of *T. cruzi* amastigote forms. However, 11-methyl cryptolepine (**50**) was about 5 times more active against *P. falciparum* and slightly more active against *T*. *cruzi* amastigote forms than the triflate salt of cryptolepine Fig. (**6**).

 Another study involving the syntheses of analogs of cryptolepine and their evaluation showed improved antiplasmodial activity as compared to the parent molecule [46]. *In vitro* studies of 2- and 7-bromocryptolepine proved to be 2-fold more potent than cryptolepine. The most potent compound revealed was 2, 7-dibromocryptolepine (**51**) with 10-fold more activity than cryptolepine (as sulfate)  $(IC_{50} =$ 0.049 and 0.44  $\mu$ M respectively against K-1 and 0.026 and 0.27 µM against HB3). It was also shown in *in vivo* studies that **51** suppressed parasitemia (*P. berghei*) by 89 % as compared to untreated infected controls at a dose of 12.5 mg kg<sup>-1</sup>.

 A more elaborate structure-bioactivity study was done on 16 cryptolepine analogs in which six of the analogs demonstrated potent *in vitro* activities against *P. falciparum* with IC<sub>50</sub> values  $\leq 0.1$  µM, and cytotoxicities 2-4 times greater than that of cryptolepine (Table **1**) [48]. Among the compounds evaluated, two novel compounds (**52** and **53**) were shown to have encouraging antimalarial activities *in vivo* with suppression of parasitemias (*P*. *berghei*) >90 % at doses of 25 mg  $kg^{-1}$  and increased survival of the mice without any apparent toxicity. According to these authors, the antimalarial mode of action in these compounds appears to be similar to that of chloroquine and may involve inhibition of hemozoin formation. It is also possible that the effect of these agents on DNA synthesis, cleavage and inhibition of topoisomerase II may contribute to their action [13d]. A recent study using fluorescence microscopy, suggests that cryptolepine accumulates in parasite structures that may correspond to the parasite nucleus [29].

 As part of our on-going studies of cryptolepine analogs, we tested several compounds against *P*. *falciparum*. One compound (**54**) showed significant potency against both D6



**Fig. (6).** Modified Indolo[3, 2-b]quinolines as Anti-Malarial and Anti-Trypanosomal agents.





and W2 clones of the organism ( $IC_{50} = 62$ , 28 ng/mL respectively) [43]. More recently, we reported the evaluation of N-10 isosteres of cryptolepine [9a]. The results demonstrated that both the sulfur ( $IC_{50} = 1.6$ , 2.0  $\mu$ g/mL) and oxygen ( $IC_{50}$  $= 0.28$ , 0.28  $\mu$ g/mL) isosteres have moderate activities against the D6 and W2 clones of *P. falciparum*. Thus, it would appear that cryptolepine and its analogs possess a new scaffold for development as antimalarial agents. Interestingly, the same compounds demonstrated potent activity against leishmaniasis *in vitro*. In fact, most of the analogs evaluated were more potent than pentamidine which served as a positive control in the evaluation.

# **C**. **Antifungal Activity**

 Opportunistic fungi have become important human pathogens and constitute a primary mode of mortality in immunocompromised patients [49]. Cryptococcus meningitis for example, is a fatal opportunistic infection if left untreated and is one of the many consequences of opportunistic infections associated with AIDS. Amphotericin B in combination with Flucytosine is the preferred treatment for this disease. However, Amphotericin B lacks oral efficacy and induces several side effects including nephrotoxicity, leukopenia, thrombocytopenia and febrile reactions. Agents from the azole class like ketoconazole and fluconazole were introduced as possible replacements for amphotericin B, but fungal resistance development has limited their broad utility and has inspired the development of new effective drug entities [49].

 Cryptolepine re-isolated from *C. sanguinolenta*, has shown potential antifungal and antimicrobial activities [9]. The reported advantages of cryptolepine over amphotericin B are its relatively low toxicity  $(LD_{50} = 146 \text{ vs } 2.3 \text{ mg/Kg})$ weight of mice) and better water solubility (1.2 *vs* 0.1 mg/mL), making it an attractive lead compound for antifungal drug development. In addition, cryptolepine shows a higher activity against *C*. *neoformans* than amphotericin B when tested in agar well diffusion assays [9b]. Several reports have now demonstrated that cryptolepine and several of its analogs have broad spectra of activities against opportunistic fungal and bacterial infections associated with AIDS [37, 38b]. A systematic study was also conducted on the quindoline skeleton to aid the formulation of SAR and the identification of potent agents with a broad spectrum of activity against various pathogens.

 Alkylation of the N-5 or/and N-10 atoms in quindoline were contemplated to understand if buckling is a necessary requirement for activity since the flat quindoline without

alkylation at N5 is devoid of activity [50]. It was found that compounds **1a** and **55** showed good activities (MIC =  $12.5$ ) and 6.25  $\mu$ g/mL) but compound **56** was only marginally active (MIC  $>62.5 \mu g/mL$ ) compared to Amphotericin B (0.39  $\mu$ g/mL) which suggests that the ionized form of cryptolepine may be the active form and that buckling is not a necessary requirement for anticryptococcal activity Fig. (**7**) [50]. Following the establishment that N-5 methylation of quindoline is a prerequisite for antifungal activity, homologation of the alkyl chain was carried out by Ablordeppey [50] to obtain the chain length necessary for optimum activity.

 In this regard, the methyl group on compound **1a** was replaced with ethyl, propyl, pentyl and octyl groups and the resulting compounds were evaluated against *C. neoformans* and *C. albicans*. There was little change in activity against both organisms through this variation. The effect of electron density on the N-5 atom was also explored. This was achieved by substituting one of the hydrogen atoms on the N-5 methyl group with electron donating or withdrawing groups to yield N-5 substituents such as  $CH<sub>2</sub>CN$ ,  $CH<sub>2</sub>OCH<sub>3</sub>$ ,  $CH<sub>2</sub>Ph$  and  $CH<sub>2</sub>COPh$ . Again, it was found that these changes resulted in little or no change in activity. A surprising observation in the activity profile of these compounds was the absence of activity associated with the N-benzylsubstituted analog. This led to the hypothesis that lack of activity may be due to a steric requirement for substituents accommodated close to the N-5 atom. An alternative hypothesis was that high electron density in the N-5 region due to the phenyl ring may be responsible for the lack of activity. Replacement of the phenyl ring with cyclohexyl and cyclopentyl rings restored activity somewhat (MIC =  $62.5$ ) mg/mL) but further reduction of the ring size (cyclobutyl and cyclopropyl) led to only minimal increase in activity. This suggests that while a flat aromatic ring is disfavored around the N-5 region of quindoline, smaller and flexible ring substituents appear to be tolerated. Extending the phenyl ring away from the N-5 atom (**57a-d**) restored activity and in fact led to significant activity increase above that of cryptolepine Fig. (**7**). The optimum chain length was found to be that associated with an  $\omega$ -phenylpentyl moiety. Subsequently, it was found that substituting a cyclohexyl ring for the phenyl ring led to a similar observation. Thus, compound **57c** was found to be about eight times more potent than cryptolepine (MIC = 1.9  $\mu$ g/mL) and constitutes the most active anticryptococcal agent in the series. Extending the chain length beyond five carbons led to a reduction in activity. Compound  $57d$  with the octyl carbon chain (MIC = 10) g/mL) is as active as cryptolepine, but five times less active than **57c**. Modifying the electron density around the N-5



**Fig. (7).** Structures of compounds **55-57**.

atom through substitution on other positions on the quindoline moiety was hypothesized as a means of modulating electron density without steric interference [51]. Position 2 was selected for evaluation because it is para to the N-5 atom and can influence the electron density around the N-5 atom through resonance effect. To this end, various electrondonating and electron-withdrawing substituents were introduced at position 2 resulting in activities against both *C. neoformans* and *C. albicans*, two important opportunistic pathogens associated with AIDS. The active analogs of cryptolepine with substitutions at position 2 are listed in Table **2**.

 However, the activity profile appears to be complex as both types of substituents enhance activity. For example, it is observed that while strong electron withdrawing groups like  $CN$  and  $NO<sub>2</sub>$  enhanced activity, substitution with sulfoxides and sulfones led to reduction in activity. Hydrophobicity did not seem to play a significant role on the SAR of these compounds. These observations suggest the possibility that multiple mechanisms of fungal inhibition may be involved.

Subsequent attempts to combine the  $\omega$ -cyclohexylpentyl or  $\omega$ -phenylpentyl group on the N-5 atom with electronwithdrawing/donating groups at position 2 and unsubstituted/methyl groups on the N-10 atom [51] resulted in a mixed effect on activity. The brominated analog (**58**) retained activity against both *C. albicans* and *C. neoformans*  (MIC = 1.9  $\mu$ g/ml for both) compared to Amphotericin B (0.39  $\mu$ g/ml). Substitution of the –NO<sub>2</sub> group at the 7 and 11 positions (**59, 60**) showed similar high potency (MIC for *C*. *neoformans* 0.25 and for *Aspergillus fumigatus* = 0.39  $\mu$ g/ml) compared to 0.39  $\mu$ g/ml for Amphotericin B Fig. (8). A tetracyclic hybrid (**61**) was synthesized integrating the pharmacophore of another antifungal natural product onychine (**62**) [52] and cryptolepine. This compound however, showed no activity against *C. albicans* and *C. neoformans*. Similarly, its N-5 methylated analog (**63**) was also inactive against *C. neoformans* and only marginally active against *C. albicans* (MIC =  $62.5 \mu g/mL$ ). The structure of the tri-cyclic onychine  $(9)$  inspired the synthesis of  $\delta$ -carbolines **64a** and **64b**. Introduction of an  $\omega$ -phenyl or  $\omega$ -cyclohexyl alkyl moieties at the N-1 position of  $\delta$ -carboline was evaluated. Here too the  $\omega$ -cyclohexylpentyl substituted analog (64a) exhibited a broad spetrum of activity (MIC in  $\mu$ g/ml were 6.25 against *C. albicans*, 0.39 against *C. neoformans*, 12.5 against *A*. *fumigatus*, 12.5 against *A. flavus* and 6.25 against M. *intracellulare*) Fig. (**9**).

 Thus, the antifungal profile of **64a** is somewhat comparable to that of amphotericin B with values at 0.2, 0.2, 0.39 and 0.39 µg/ml against *C. albicans, C. neoformans, A. fumi*gatus and A. flavus respectively. Similar evaluation of  $\omega$ -

# **Table 2. Effect of Substitutions at Position 2 on the** *In Vitro* **Antifungal Activity [51]**





 $MIC = Minimum Inhibitory Concentration, Cn = C, neoformans; Ca = C, albicans$ 

 $Af = A.$  fumigatus;  $NT = Not$  tested.



**Fig. (9).** Structures of compounds **61-64**.

phenylpentyl-substituted analog (**64b**) showed a more subdued activity against *C. albicans, C. neoformans* and *A. fu* $migatus$  (MIC = 31.2, 0.98 and 125  $\mu$ g/ml respectively) [52]. Further evaluation using various combinations of arylalkyl substituents on N-5 and N-10 atoms of the indoloquinoline ring was not very productive [43].

 Synthesis and evaluation of doubly methylated (N-5 and N-10) analogs (**55, 65a, 65b**) showed that substitution at the 2-position with the electron withdrawing bromine (**65a**) while enhancing anticryptococcal activity [52] has little action on anticandida acitivity Fig. (**10**). Similarly, the electron-donating methoxy group at the 2-position (**65b**) has little or no effect on either activity when compared to **55**. It was also reported that alkylation of the N-10 atom with  $\omega$ cyclohexylpentyl moiety without alkylation on N-5 resulted in no activity against *C. neoformans* and *C. albicans* [43]. Since simultaneous alkylation of both N-5 and N-10 appears to be useful, bis-alkylated compounds (**66a-c**) were synthesized and evaluated for activity. The analog with the  $\omega$ phenylpentyl moiety on N-10 and methyl group on N-5 (**66a**) turned out to be more potent than the reversed substitution against *C. neoformans* and *C. albicans* (66a,  $IC_{50} = 2.5 \& 2.0$ g/ml respectively) [43]. Bis-quindolinium derivatives with an alkyl bridge connecting N5/N5 (**67a-b**) and those connecting N10/N10 atoms of the quindolines (**68**) were also synthesized and evaluated Fig. (**10**). It was observed that a positively charged quaternary nitrogen is also required for







**Fig. (10).** Structures of compounds **65-68**.



**66a** R = H;  $R_5$  = CH<sub>3</sub>;  $R_{10}$  = (CH<sub>2</sub>)<sub>5</sub>-C<sub>6</sub>H<sub>6</sub>; X = I **66b** R = H;  $R_5 = (CH_2)_5-C_6H_5$ ;  $R_{10} = CH_3$ ;  $X = Br$ **66c** R = Br;  $R_5 = (CH_2)_5 - C_6H_5$ ;  $R_{10} = CH_3 X = Br$ 



activity in bis-quindolinium compounds. Also noted was the fact that a bis-quindolinium compound separated by n-pentyl carbon bridge (**67b**) was more potent than the corresponding analog separated by four carbon atoms (**67a**). A similar observation was made about the N-10/N-10 five-carbon connected bis-quindolinium compound (**68**) [43].

 At the beginning of 2007, we [9a] reported the effect of isosteric replacement of the N-10 in cryptolepine on antifungal activity. It was shown that antifungal activity diminished when the N-10 was replaced by oxygen and carbon but activity was retained when the replacement was sulfur. Interestingly, the benzothienoquinoline (sulfur substituents at position 10) analog of cryptolepine showed a broad spectrum of activity and was significantly less toxic than the parent compound. Thus, the authors suggested that the benzothieno[3,2 b]quinolines might serve as a new scaffold for the development of antifungal drugs.

### **D**. **Renal Vasodilation and Inhibition of Platelet Aggregation**

 The need for new drugs for cardiovascular diseases is underscored by this statement from the World Health Organization: "Cardiovascular disease is the number one cause of death globally and is projected to remain the leading cause of death. An estimated 17.5 million people died from cardiovascular disease in 2005, representing 30 % of all global deaths. Of these deaths, 7.6 million were due to heart attacks and 5.7 million were due to stroke. Around 80% of these deaths occurred in low and middle income countries (LMIC). If appropriate action is not taken, by 2015, an estimated 20 million people will die from cardiovascular disease every year, mainly from heart attacks and strokes." [53].

 Much of the earlier biological investigations on cryptolepine were related to its effect on the cardiovascular system. The effect of cryptolepine on renal vasodilation was reported by Oyekan in 1994 [11]. This effect in rats appears to be endothelium-dependent and involves the release of a noncyclooxygenase endothelium-dependent relaxing factor (EDRF), possibly nitric oxide (NO). The author further reported that the effector pathway for cryptolepine-induced vasodilation is by cGMP of vascular smooth muscle. In addition, the vasodilatory effect was proposed to be mediated *via* K<sup>+</sup> efflux, notably through activation of tetraethyl ammonium-sensitive but glibenclamide-insenstive K+ channels and Na+/K+-ATPase. Here, unlike diazoxide and acetylcholine, the  $K^+$  channels activated by cryptolepine are not coupled to pertussis toxin-sensitive G-proteins. However, many interwoven mechanisms seem to be involved which are directly related to the central mechanism involving the release of EDRF [54]. Cryptolepine inhibits platelet aggregation *in vitro* and *in vivo* and stimulates fibrinolysis *ex vivo* [10a]. The mechanism appears to involve an increase in platelet cAMP levels, possibly through stimulation of platelet adenylate cyclase [10c]. The antiaggregatory effects do not seem to invovlve extracellular or intracellular  $Ca^{2+}$ . Cryptolepine also inhibits collagen-induced secondary aggregation through a selective antiphospholipase-like activity. However, there was no effect on platelet cyclooxygenase and lipoxygenase activities of platelets [10d]. A limited structure activity relationship was done around the N-5 position of the quindoline ring in inhibiting ADP-induced aggregation and renal vasodilation [55]. Apart from quindoline, 4 analogs were synthesized and tested of which 5-ethyl quindoline was the most potent while quindoline the parent compound, was without effect. In fact, the 5-ethylquindoline was 2.5 times more potent than cryptolepine. The rank order of renal vasodilator and antiplatelet effect following alkyl substitution is: ethyl>methyl>propyl>benzyl. This trend suggests that the N-5 position has limited bulk tolerance and lipophilicity with an ethyl group being the optimum alkyl substituent. Although it was suggested that distortion of the flat quindoline structure due to alkylation at N-5 position may explain the enhanced activity, more recent observations on the antifungal properties may suggest instead the development of the positive charge on N-5 may be a more important factor in the expression of these activities.

### **E**. **Anti Hyperglycemic Activity**

 Hyperglycemia or high level of glucose in blood is one of the classic symptoms of diabetes (I or II) and left untreated can lead to organ damage. People who are older, overweight, have family members with diabetes, or are from certain ethnic groups are at greater risk for developing hyperglycemia [56a]. Hyperglycemia can be caused by insulin resistance, impaired glucose metabolism, medications or the use of some nutritional supplements. Several plants and plant extracts are known to have antihyperglycemic properties and a number of crude plant extracts and purified substances from plants have been under clinical trials. [56b-d]

 Bierer and coworkers [20] were the first to report the antihyperglycemic properties of cryptolepine. They have synthesized several salts including the hydroiodide, hydrochloride and hydrotriflate of cryptolepine (**1a, 1b** and **1d**) and tested for their ability to stimulate glucose transport in 3T3-L1 adipocytes. It was observed that cryptolepine showed an increasing ability to stimulate glucose transport beginning at 3 µM. Cryptolepine hydrochloride lowered blood glucose in Non-Insulin-Dependent Diabetes Mellitus (NIDDM) animal models with reduced food intake. Fasting time course studies and a pair-fed study in db/ab mice seem to indicate that this antihyperglycemic effect is separate from the anorexigenic effect. The presence of the methyl substituent at N-5 appears to be critical for activity as the N-10 methyl regioisomer was inactive *in vitro* and *in vivo*. Cryptolepine hydrochloride lowered serum glucose concentrations in fructose-fed STZ-treated rat model of NIDDM by 16 %, 34 % and 45 % on days 1, 2 and 3 of dosing respectively. The compound also lowered serum triglyceride concentrations by 30 %, 39 % and 69 % on days 1, 2 and 3 of dosing respectively. There was a significant decline in food consumption (29 %,  $p < 0.05$ ) however, the body weight gain over the course of the experiment was not altered.

 In a preliminary structure-antihyperglycemic activity studies conducted on cryptolepine reported by the same authors [27], a series of substituted cryptolepine analogues were synthesized and their bioactivities were measured in a 3T3-L1 glucose transport assay. Fig. (**11**). A group of these compounds was further evaluated in genetically altered obese diabetic C57BL/KS-db/ab mice. The results indicated that two derivatives (**1b** and **69**) out of 33 analogs for which SAR



**Fig. (11).** Analogs of Cryptolepine tested for antihyperglycemic Activity.

studies were conducted, have true antihyperglycemic activity. *In vitro* studies showed that increasing the lipophilicity of the substituent at N-5 increased the glucose transport activity. A chloro substitution at the C-11 position of the ring reduced the activity of the cryptolepine analog possibly due to a deactivating effect. However, simultaneous substitution of a chloro atom at positions 1 and 11 (**70**) showed similar activity as the parent compound (**1b**). Substitution at the 4 position with OMe, (**69**) or at the 6-position with fluorine (**71**) improved activity, possibly due to an interaction between the methoxy/fluoro group and the N-5 methyl group, altering the planarity of the molecule. Replacement of the nitrogen at the 10-position with oxygen, sulfur and methylene groups led to a reduction in the glucose transport activity. Later, *in vivo* studies showed that the methoxy derivative (**69**) was the most efficacious derivative tested, lowering the plasma glucose level by 58.3 %, at 24 h post dose while showing improved food intake as compared with cryptolepine hydrochloride. The 2-fluoro derivative (**72**) was also efficacious, lowering the plasma glucose level by 42.5 % at 2 h post dose, with activity still being observed at 24 h. Interestingly, compound **72** showed no signs of toxicity at higher concentrations (30 mM). However, as with **1b**, this effect was accompanied by a significant reduction in both mean body weight and food intake. Substitution at position 11 decreased the ability of the compound to lower plasma glucose. This observation is similar to the SAR observed for antifungal activity and suggests similarity in the mode of action in these biological actions. It was also observed that compound **73,** with an 11-anilino and 2-fluoro groups, showed improvement in food consumption and body weight. Increasing the steric bulk at the N-5 position as in compounds **74** and **75** did not change their ability to lower plasma glucose but produced an improvement in food intake and body weight parameters. The benzyl derivative (**74**) was observed to have lowered plasma glucose by 15 % at 3 h post dose with no

effect on food intake or body weight while the 4-fluorobenzyl derivative (**75**) lowered plasma glucose by 36.3 % with improved food intake. This appears to suggest that a fluoro substitution has a beneficial effect on lowering plasma glucose levels in a similar manner irrespective of its position on the scaffold [*cf.* compound **72**, Fig. (**11**)].

#### **F**. **Anticancer Activity and Cytotoxicity**

 According to the World Health Organization (WHO) reports, more than 11 million people are diagnosed with cancer every year and it is estimated that there will be 16 million new cases every year by 2020. Cancer causes 7 million deaths every year or 12.5% of deaths worldwide. Plant derived drugs such as etoposide, taxol, vincristine and several others have contributed to current treatment modalities. Despite recent advances of cancer chemotherapy, the cytotoxic effects of the present drugs on non-cancerous cells have not been addressed satisfactorily. The need for new drugs is therefore indicated and plant sources continue to be important avenues of new drug discovery.

 Cryptolepine has been shown to possess cytotoxic activity (IC<sub>50</sub> = 1.3  $\mu$ M) through the inhibition of DNA synthesis. Its cytotoxic potency is estimated to be 4-5 times more than that of ellipticine towards B16 melanoma cells and therefore a valid candidate for the development of anti-tumor agents [13a]. It easily crosses cell membranes and accumulates selectively in nuclei rather than in the cytoplasm of B16 melanoma cells and blocks the cell cycle in G2/M phase. However, in human lung adenocarcinoma A549 cells, the cell cycle arrest was at either G1 phase or G2/M phase, depending on the concentration [13b]. In addition to its interaction with DNA, cryptolepine is reported to target topoisomerase II in susceptible cells. Cryptolepine binds tightly to DNA and behaves as a typical intercalating agent [18]. The binding affinity is similar to that of other intercalators such as

actinomycin D, daunomycin, porphyrin compounds and chromomycin. Specifically, cryptolepine prefers to bind to C-G rich sequences of DNA, with a tendency for nonalternating CC sites. A recent crystal structure of cryptolepine-d(CCTAGG) complex confirms the same [58]. Dassonneville *et.al* [59] provided direct evidence that cryptolepine induces apoptosis in HL-60 leukemia cells ( $IC_{50} = 3.2 \mu M$ ) and is two times less toxic to HL-60/MX2 cells resistant to the antitumor drug mitoxanthrone. According to these authors, mitochondria and caspases play a central role in the activation of the executioner phase of cryptolepine induced apoptosis.

 Cryptolepine also exhibited weak telomerase inhibitory activity  $($ <sup>Tel</sup>IC<sub>50</sub> 9.3  $\mu$ M). However, there is moderate PCR inhibition at  $\sim$ 5  $\mu$ M, suggesting a low degree of selectivity for quadruplex over duplex DNA. It is also suggested that cryptolepine stabilizes topoisomerase II-DNA covalent complex and stimulates the cutting of DNA by topoisorerase II [60]. It is believed that cryptolepine produces cell death sig-

#### *Indolo[3,2-b]quinolines: Synthesis, Biology and SAR Mini-Reviews in Medicinal Chemistry,* **2008***, Vol. 8, No. 6* **551**

nals other than topoisomerase II-mediated DNA breakage and topoisomerase II inhibition may play a minor role in the cytotoxicity of cryptolepine. As with most of the other anticancer agents, the mechanism of action of cryptolepine is probably pleiotropic.

#### *a) Indoloquinoline Derivatives as DNA Intercalators*

 Several cryptolepine derivatives based on modifications at C-2, N-5, N-10 and C-11 positions were synthesized and their structure activity relationship towards cytotoxicity studied Fig. (**3**) [61]. Initially, all the compounds were tested on 1138 yeast strain and selected analogs (**1d, 76-81**) were tested for cytotoxicity to M109 Madison lung carcinoma cell culture Fig. (**12**). The most active compounds tested for cytotoxicity against M109 Madison lung carcinoma cell culture were cryptolepine hydrotriflate (**1d**) and the N-acetyl derivative (**78**). Compounds **1d** and **78** were further subjected to analysis of their effect on cell cycle dependent growth disturbances. The results showed a G1 cell cycle arrest and apoptosis in A2780/p53wt cells at 5 x  $IC_{50}$ . More recently,



**Fig. (12).** Indoloquinoline analogs as DNA intercalators.

Matsui [13d] and colleagues reported that cryptolepine induces the expression of p21WAF1/CIP1 with growth arrest in p53-mutated human osteocarcinoma MG63 cells. Interestingly, their results showed that cryptolepine inhibited the growth of MG63 cells and caused a G2/M-phase arrest.

 Higher concentrations showed a dose-dependent increase in apoptosis. Chen and co-workers [62, 63] studied cell growth inhibition for a series of substituted indoloquinoline-6-carboxamides and evaluated their activity in a panel of cell lines including Murine P388 leukemia, Murine Lewis lung carcinoma and Human Jurkat leukemia. Substitution of indolo[1,2-b]quinoline-6-carboxamides (**82a** and **82b**) and 11H-benzothieno[3, 2-b]quinoline-4-carboxamides (**83a** and **83b**) at the pseudo-peri position to the carboxamide side chain provided analogs of significantly increased cytotoxicity;  $IC_{50} = 450, 77, 170$  and 171 nM respectively on Human Jarkat leukemia  $J_{c}$  as compared to the dual topo I/II inhibitor, N- [2- dimethyl-amino) ethyl]acridine-4-carboxamide (DACA) with an  $IC_{50}$  of 580 nM. The monomeric quindolines (**82** and **84**) discriminate poorly between cells with varying levels of topoisomerase II. The carboxamide-linked bis(quindoline) (**85a**) was less potent than **84a** in P388, but more potent in the human leukemia cell lines  $(IC_{50} = 24 \text{ vs } 100 \text{ m})$ 230 nM) Fig. (**12**). The corresponding bis(4-methylquindoline) (**85b**) had a broadly similar pattern of cytotoxicity. The bis- analogs (**86a** and **86b**) displayed poor activity in P388 and Lewis lung lines, but **86b** was unexpectedly more active in the human leukemia cell lines ( $IC_{50} = 4.5$  nM). In contrast to the monomers, the cytotoxicities of the dimeric quindolines are inversely proportional to the level of topoisomerase II in the cells. Compound **86c** was nontoxic  $(IC_{50} = 9900$ nM) while compound **86d** was among the most active analogues ( $IC_{50} = 13.9$  nM). In all the cases, DACA was used as the standard with an  $IC_{50}$  value of 580 nM. The  $IC_{50}$  ratios of  $J_{A}/J_{C}$  for the above compounds were less than about 2-fold suggesting a non-topo II mediated mechanism while DACA

has the ratio of about 2.0 and behaves as a topo I/II inhibitor. Wright and coworkers [46] identified compound **87** with 4-5 fold more cytotoxicity to MAC15A cells than cryptolepine hydroiodide. Mardenborough [43] synthesized dimeric cryptolepine analogs (**45** and **46**) Fig. (**5**), which displayed no cytotoxicity up to  $23.8 \mu g/mL$  on Vero cells, suggesting the dimeric compounds to be less toxic than their monomeric counterparts.

 One concern regarding the use of cryptolepine analogs as anticancer agents is the lack of selectivity between DNA intercalation of cancer and normal cells. A solution to this problem has recently been proposed by Wright and colleagues at the University of Bradford. They proposed the development of bioreducible prodrugs that would be bioreductively activated by nitroquinone oxido-reductases present in tumor cells at higher concentrations than in normal cells [13e]. With a limited number of nitrocryptolepine derivatives, the authors were able to demonstrate that some of the compounds are cytotoxic and may be substrates for the oxido-reductases (NOO1  $\&$  2). Unfortunately, the compounds did not show significant toxicity towards the cells with high NQO1 (H460 cells) compared to those with low NQO1 (BE cells).

# **b**) **Telomerase II Inhibition**

 Folding of telomeric DNA into a four-stranded structure involving G-quartets has been shown to inhibit enzymes from catalyzing the synthesis of telomeric DNA repeats. Telomerase is expressed in tumor cells, but not in most somatic cells. Synthetic small molecules that induce and stabilize such G-quadruplex structures are potential telomerase inhibitors and could interfere with telomere elongation and replication of cancer cells. Several quindoline derivatives were designed and synthesized by Zhou and coworkers [64] and their interaction with the telomeric G-quadruplex has been studied Fig. (**13**).



**Fig. (13).** Cryptolepine/Indoloquinoline analogs as Telomerase inhibitors.

 Introducing electron-donating groups such as a substituted amino group at position 11 of the quindoline scaffold significantly increased the electron density at the N-5 atom, resulting in easy protonation under physiological pH. EMSA and CD spectral studies showed that these compounds were capable of inducing the formation and stabilization of the Gquadruplex and thus exhibited inhibitory effect on telomerase. Competition dialysis experiments showed that compound **88** interacted preferentially with G-quadruplex and had comparatively weak affinity for duplex DNA. *In vitro* inhibitory activity studies on these compounds were done using telomerase repeat amplification protocol (TRAP) assay. The results showed that telomerase inhibition properties of the quindoline derivatives under study were significantly improved upon introduction of electron donors into the parent compound. Compounds **88** and **89** (with strong electron donor substituents in the 11 position) showed better  $^{Tel}IC_{50}$ values  $(0.55\mu M$  and  $0.44 \mu M$  respectively) as compared to quindoline  $(>138 \mu M)$  in that series.

 Another report by Capiro [65] observed that **3** has significant activity against human telomerase  $({}^{Tel}IC_{50} 16 \mu M,$  as compared to 2, 6-dimethylamino-anthraquinone, 4.1  $\mu$ M), a level comparable to several nucleoside inhibitors of reverse transcriptase activity. Compound **90** is approximately 10 fold less active than the best quadruplex-mediated inhibitors based on tricyclic chromophores and is 2-fold less active than the standard compound (91) ( $^{Tel}IC_{50}$  7.0  $\mu$ M). More recently, Guyen [66] reported 2, 7-dialkylamine substituted quindolines (92 and 93) have improved potency  $(^{Tel}EC_{50}$  values of 6.3  $\mu$ M and 11.8  $\mu$ M) and did not exhibit any noticeable PCR inhibition at up to 50  $\mu$ M Fig (4). This is suggestive of selectivity for quadruplex over duplex DNA Fig. (**13**).

### **CONCLUSION**

 Indolo[2,3-b]quinolines constitute a lead scaffold for drug development. Synthetic strategies for obtaining the tetracyclic structure are now available providing flexibility for introducing various functional groups into the ring system. Such functionalized structures are associated with a variety of pharmacological activities including antiinfective, vasodilation, antiplatelet aggregation, antihyperglycemic, anticancer and several others. Structure activity relationship studies are beginning to emerge. However, a consistent mechanism of action by which the tetracyclic ring is involved in the various biological actions is yet to emerge. Clearly, this is one area of focus for the future.

### **ACKNOWLEDGEMENT**

 The authors gratefully acknowledge the editorial contributions of Mrs. Barbara Bricker. This work would not have been possible without the continuing financial support of RCMI grant number G12 RR 03020 from NCRR and the Title III grant to SYA.

# **REFERENCES**

- [1] a) Fitchter, F.; Boehringer, R. *Chem. Ber*., **1906**, *39*, 3932; b) Fitchter, F.; Probst, H. *Chem. Ber*., **1907**, *40*, 3478; c) Fichter, F.; Rohner, F. *Chem. Ber*., **1910**, *43*, 3489.
- [2] Clinquart, E. *Bull. Acad. Med. Belg*., **1929**, *9*, 627.
- [3] a) Delvaux, E. *J. Pharm. Belg*., **1931**, *13*, 955; b) Delvaux, E. *J. Pharm. Belg*., **1931**, *13*, 973.

#### *Indolo[3,2-b]quinolines: Synthesis, Biology and SAR Mini-Reviews in Medicinal Chemistry,* **2008***, Vol. 8, No. 6* **553**

- [4] Gunatilaka, A. A. L.; Sotheeswaran, S.; Balasubramaniam, S.; Chandrasekara, A. I.; Sriyani, H. T. B. *Planta Med*., **1980**, *39*, 66.
- [5] Gillert, E. *Helv. Chim. Acta,* **1951**, *34*, 642.
- [6] a) Dwuma-Badu, D.; Ayim, J. S. K.; Fiagbe, F. N. I. Y.; Knapp, J. E.; Schiff, P. L., Jr.; Slatkin, D. J. *J. Pharm. Sci*., **1978**, *67*, 433; b) Ablordeppey, S. Y.; Hufford, C. D.; Borne, R. F.; Dwuma-Badu, D. *Planta Med.,* **1990**, *56*, 416.
- [7] a) Cimanga, K.; Pieters, L.; Claeys, M; Vanden Berghe, D.; Vlietinck, A. J. *Planta Med*., **1991**, *57* (Suppl. 2), A98; b) Paulo, A.; Pimentel, M. Viegas, S.; Pires, I.; Cabrita, J.; Gomes, E. T. *J. Ethnopharmacol.,* **1994**, *44*, 73; c) Paulo, A.; Duarte, A.; Gomes, E. T. *J. Ethnopharmacol.,* **1994**, *44*, 127.
- [8] a) Wright, C. W. *J. Pharm. Pharmacol.,* **2007**, *59*, 899; b) Noamesi, B. K.; Paine, A.; Kirby, G. C.; Warhurst, D. C.; Phillipson, J. D. *Trans. R. Soc. Trop. Med. Hyg.,* **1991**, *85*, 315; c) Kirby, G. C.; Noamesi, B. K.; Paine, A.; Warhurst, D. C.; Phillipson, J. D. *Phytother. Res*., **1995**, *9*, 359; d) Cimanga, K.; De Bruyne, T.; Lasure, A.; Van poel, B.; Pieters, L.; Claeys, M; Vanden Berghe, D.; Kambu, K.; Tona, L.; Vlietinck, A. J. *Planta Med.*, **1996**, *62*, 22; e) Wright, C. W.; Phillipson, J. D.; Awe, S. O.; Kirby, G. C.; Warhurst, D. C.;Quetin-Leclercq, J.; Angenot, L. *Phytother. Res.,* **1996**, *10*, 361; f) Cimanga, K.; De Bruyne, T; Pieters, L.; Vlietinck, A. J.; Turger, C. A. J. *Nat. Prod*., **1977**, *60*, 688.
- [9] a) Zhu, X. Y; Mardenborough, L. G.; Li, S.; Khan, A.; Zhang, W.; Fan, P.; Jacob, M.; Khan, S.; Walker, L.; Ablordeppey, S. Y. *Bioorg. Med. Chem*., **2007**, *15*, 686; b) Singh, M.; Singh, M. P.; Ablordeppey, S. Y. *Drug Develop. Indust. Pharm*., **1996**, *22*, 377.
- [10] a) Oyekan, A. O.; Botting, J. H.; Noamesi, B. K. *Gen. Pharmacol*., **1988**, *19*, 233; b) Oyekan, A. O.; Okafor, J. P. *J. Ethnopharmacol*., **1989**, *27*, 141; c) Oyekan, A. O.; Ablordeppey, S. Y. A. *Gen. Pharmacol*., **1993**, *24*, 461; d) Oyekan, A. O.; Ablordeppey, S. Y. A. *Gen. Pharmacol*., **1993**, *24*, 1285.
- [11] Oyekan, A. O. *J. Cardiovasc. Pharmacol*., **1994**, *23*, 602.
- Luo, J.; Fort, D. M.; Caarlson, T. J.; Noamesi, B. K.; Amon-Kotei, D.; King, S. R.; Tsai, J.; Quan, J.; Hobensack, C.; Lapresca, P.; Waldeck, N.; Mendez, C. D.; Jolad, S. D.; Bierer, D. E.; Reaven, G. M. *Diabet. Med*., **1998,** *15*, 367.
- [13] a) Bonjean, K.; De Pauw-Gillet, M. C.; Defresne, M.-P.; Colson, P.; Houssier, C.;.Dassonneville, L.; Bailly, C.; Wright, C.; Quetin-Leclercq, J.; Angenot, L. *Anticancer Res*., **1997**, *17*, 4068; b) Dassonneville, L Bonjean, K.; De Pauw-Gillet, M. -C.; Colson, P.; Houssier, C.; Quetin-Leclercq, J.; Angenot, L.; Bailly, C. *Biochemistry,* **1999**, *38*, 7719.; c) Ansah, C.; Gooderham, N. J*. Toxicol. Sci*., **2002**, *70*, 245; d) Matsui, T-A.; Sowa, Y.; Murata, H.; Takagi, K.; Nakanishi, R.; Aoki, S.; Yoshikawa, M.; Kobayashi, M.; Sakabe, T.; Koichi, T.; Sakai, T. *Int. J. Oncol.,* **2007**, *31*, 915; e). Seville, S.; Phillips, R. M.; Shnyder, S. D.; Wright, C. W. *Bioorg. Med. Chem.,* **2007**, *15*, 6353.
- [14] a) Bamgbose, S. O.; Noamesi, B. K. *Planta Med*., **1981,** *41*, 392; b) Noamesi, B. K.; Bamgbose, S. O. *Planta Med*., **1984**, *44*, 98; c) Olajide, O. A.; Heiss, E. H.; Schachner, D.; Wright, C. W.; Vollmar, A. M.; Dirsch, V. M. *Bioorg. Med. Chem*., **2007**, *15*, 43.
- [15] a) Hamet-R. *C. R. Soc. Biol*., **1937**, *126*, 768; b) Hamet-R. *Acad. Sci. Belg*., **1938**, *207*, 1016; c) Noamesi, B. K.; Bamgbose, S. O. A. *Planta. Med*., **1980**, *39*, 51.
- [16] Noamesi, B. K.; Bamgbose, S. O. A. *Planta Med*., **1982**, *44*, 241.
- [17] Rauwald, H. W.; Kober, M.; Mutschler, E.; Lambrecht, G. *Planta Med*., **1992**, *58*, 486.
- [18] Bonjean, K.; De Pauw-Gillet, M. C.; Defresne, M.-P.; Colson, P.; Houssier, C.; Dassonneville, L.; Bailly, C.; Greimers, R.; Wright, C.; Quetin-Leclercq, J.; Tits, M.; Angenot, L. *Biochemistry,* **1998**, *37*, 5136.
- [19] a) Holt, S. J.; Petrow, V. *Proc. Roy. Soc. B.,* **1946**, *148*, 607; b) Holt, S. J.; Petrow, V. *J. Chem. Soc*., **1947**, 607; c) Holt, S. J.; Petrow, V. *J. Chem. Soc.,* **1948**, 919.
- [20] Bierer, D. E.; Fort, D. M.; Mendez, C. D.; Luo, J.; Imbach, P. A.; Dubenko, L. G.; Jolad, S. D.; Gerber, R. E.; Litvak, J.; Lu, Qing.; Zhang, P.; Reed, M. J.; Waldeck, N.; Bruening, R. C.; Noamesi, B. K.; Hector, R. F.; Carlson, T. J.; King, S. R. *J. Med. Chem.,* **1998**, *41*, 894.
- [21] Fan, P.; Ablordeppey, S. Y. *J. Heterocycl. Chem*., **1997**, *34*, 1789.
- [22] Dhanabal, T.; Sangeetha R.; Mohan, P. S. *Tetrahderon,* **2006**, *62*, 6258.
- [23] Cooper, M. M.; Lovell, J. M.; Joule, J. A. *Tetrahedron Lett.*, **1996**, *37*, 4283.

#### **554** *Mini-Reviews in Medicinal Chemistry,* **2008***, Vol. 8, No. 6 Kumar et al.*

- [24] Ho, T-L; Jou, D-G. *Helv. Chim. Acta,* **2002**, *85*, 3823.
- [25] Radl, S.; Konvicka, P.; Vachal, P. *J. Heterocycl. Chem*., **2000**, *37*, 855.
- [26] Dutta, B.; Some, S.; Ray, J. K. *Tetrahderon Lett.*, **2006**, *47*, 377.
- [27] Bierer, D. E.; Dubenko, L. G.; Zhang, P.; Lu, Qing.; Imbach, P. A.; Garofalo, A. W.; Phuan, P-W.; Fort, D. M.; Litvak, J.; Gerber, E.; Sloan, B.; Luo, J.; Cooer, R.; Reaven, G.M. *J. Med. Chem*., **1998**, *41*, 2754.
- [28] Arzel, E,; Rocca, P.; Marsais, F.; Godard, A.; Queguiner, G. *Tetrahedron Lett*., **1998**, *39*, 6465.
- [29] Arzel, E.; Rocca, P.; Grellier, P.; Labaeid, M.; Frappier, F.; Gueritte, F.; Gaspard, C.; Marsais, F.; Godard, A.; Queguiner, G. *J. Med. Chem*., **2001**, *44*, 949.
- [30] Yamato, M.; Takeuchi, Y.; Chang, M.; Hashigaki, K.; Tsuruo, T.; Tashiro, T.; Tsukagoshi, S*. Chem. Pharm. Bull.,* **1990**, *38*, 3048.
- [31] Yamato, M.; Takeuchi, Y.; Chang, M.; Hashigaki, K. *Chem. Pharm. Bull.,* **1992**, *40*, 528.
- [32] Gorlitzer, K.; Weber, J. *Arch. Pharm. (Weinheim),* **1981**, *314*, 852.
- [33] Chang, M.; Takeuchi, Y.; Hashigaki, K.; Yamato, M. *Heterocycles,* **1992**, *33*, 147.
- [34] Walsh, C.; Wright, G. *Chem. Rev*., **2005**, *105*, 391.
- [35] Brown, E. D.; Wright, G. D. *Chem. Rev*., **2005**, *105*, 759.
- [36] Gibbons, S., *Nat. Prod. Rep*., **2004**, *21*, 263.
- [37] Boakye-Yiadom, K.; Heman-Ackah, S. M. *J. Pharm. Sci.*, **1979**, *68*, 1510.
- [38] a) Sawer, I.K.; Berry, M. I.; Brown, M. W.; Ford, J. L. *J. Pharm. Pharmacol.*, **1993**, *45*, 1108; b) Cimanga, K.; De Bruyne, t.; Lasure, A.; Van poel, B.; Pieters, L.; Claeys, M; Vanden Berghe, D.; Kambu, K.; Tona, L.; Vlietinck, A. *Planta Med.*, **1996**, *62*, 22.
- [39] Sawer, I. K.; Berry, M. I.; Ford, J. L. *Lett. Appl. Microbiol.,* **2005**, *40*, 24.
- [40] Sawer, I. K.; Berry, M. I.; Brown, M. W.; Ford, J. L. *J. Appl. Bacteriol.,* **1995**, *79*, 314.
- [41] Sawer, I. K.; Berry, M. I.; Ford, J. L. *Lett. Appl. Microbiol.,* **1997**, *25*, 207.
- [42] Gibbons, S.; Hallah, F.; Wright, C. W. *Phytother. Res*., **2003**, *17*, 434.
- Mardenborough, L. G.; Zhu, X. Y.; Fan, P.; Jacob, M. R.; Khan, S. I.; Walker, L. A.; Ablordeppey, S. Y. *Bioorg. Med. Chem*., **2005**, *13*, 3955.
- [44] Trape, J-F.; Pison, G.; Speigel, A.; Enel, C, Rogier, C. *Trends Parasitol*., **2002**, *18*, 224.
- [45] Wright, C. W. *Phytochem. Rev.*, **2005**, *4,* 55.
- [46] Wright, C. W.; Addae-Kyereme, J.; Breen, A. G.; Brown, J. E.; Cox, M. F.; Croft, S. L.; Y.; Kendrick, H.; Philips, R. M.; Pollet, P. L. *J. Med. Chem*., **2001**, *44*, 3187.
- [47] Grellier, P.; Ramiaramanana, L.; Millerioux, V.; Deharo, E.; Schrevel, J.; Frappier, F.; Trigalo, F.; Bodo, B.; Pousset, J-L. *Phytother. Res.,* **1996**, *10*, 317.

**Received: 05 November**, **2007 Revised: 04 December, 2007 Accepted: 05 December, 2007**

- [48] Onyeibor,O.; Croft, S. L.; Dodson, H. I.; Feiz-Haddad, M.; Kendrick, H.; Millington, N. J.; Parapini, S.; Phillips, R. M.;Seville, S.; Snyder, S.D.; Taramelli, D.; Wright. W. W. *J. Med. Chem*., **2005**, *48*, 2701.
- [49] Ablordeppey, S. Y.; Fan, P.; Ablordeppey, J. H.; Mardenborough, L*. Curr. Med. Chem.,* **1999**, *6*, 1151.
- [50] Ablordeppey, S. Y.; Fan, P.; Clark, A. M.; Alison, N. *Biorg. Med. Chem*., **1999**, *7*, 343.
- [51] Ablordeppey, S. Y.; Fan, P.; Li, S.; Clark, A. M.; Hufford, C. D. *Biorg. Med. Chem*., **2002**, *10*, 1337.
- [52] Mardenborough, L. G.; Fan, P.C.; Ablordeppey, S. Y. *Med. Chem. Res*., **1999**, *92*, 118.
- [53] http://www.who.int/cardiovascular\_diseases/en/
- [54] Oyekan, A. O. *Eur. J. Pharmacol*., **1995**, *285*, 1.
- [55] Oyekan, A. O.; Ablordeppey, S. Y. A. *Med. Chem. Res*., **1996**, 602.
- [56] a) Must, A.; Spadano, J.; Coakley, E. H.; Field, A. E.; Colditz, G.; Dietz, W. H. *J. Am. Med. Assoc.*, **1999**, *282*,1523; b) Anhauser, M. *Drug Discov. Today*, **2003**, *8*, 868; c) Atta Ur, R.; Zaman, K. *J. Ethnopharmacol*., **1989**, *26,* 1; ) Izzo, A. A.; Ernst, E. *Drugs,* **2001**, *61*, 2163.
- [57] Zhu, H.; Gooderham, N. *J. Tox. Sci*., **2006**, *91,* 132.
- [58] a) Lisgarten, J. N.; Coll, M.; Portugal, J.; Wright, C. W.; Aymami, J. *Nat. Struct. Biol*., **2002**, *9*, 57. b) Lisgarten, J. N.; Pous, J.; Coll, M.; Wright, C. W.; Aymami, J. *Acta Crystallogr. D: Biol. Crystallogr.*, **2002**, *58*, 312-313.
- [59] Dassonneville, L.; Lansiaux, A.; Wattelet, A.; Wattez, N.; Mahieu, C.; Miert, S. V.; Pieters, L.; Bailly, C*. Eur. J. Pharmacol*., **2000**, *409*, 9.
- [60] Guittat, L.; Alberti, P.; Rosu, F.; Miert, S. V.; Thetiot, E.; Pieters, L.; Gabelica, V.; De Pauw, E.; Ottaviani, A.; Riou, J. -F.; Mergny, J. -L. *Biochimie,* **2003**, *85*, 535.
- [61] Yang, S.; Abdel-kader, M.; Malone, S.; Werkhoven, M. C. M.; Wisse, J. H.; Bursuker, I.; Neddermann, K.; Fairchild, C.; Raventos-Suarez, C.; Menendez, A. T.; Lane, K.; Kingston, D. G. I. *J. Nat. Prod*., **1999**, *62*, 976.
- [62] Chen, J.; Deady, L. W.; Desneves, J.; Kaye, A. J.; Finlay, G. J.; Baguley, B. C.; Denny, W. A. *Biorg. Med. Chem*., **2000**, *8*, 2461.
- [63] Chen, J.; Deady, L. W.; Kaye, A. J.; Finlay, G. J.; Baguley, B. C.; Denny, W. A.; *Biorg. Med. Chem*., **2002**, *10*, 2381.
- [64] Zhou, J-L.; Lu, Y-J.; Ou, T-M.; Zhou, J-M.; Huang, Z-S.; Zhu, X-F.; Du, C-J.; Bu, X-Z.; Ma, L.; Gu, L-Q.; Li. Y-M.; Chan, A. S-C. *J. Med. Chem.,* **2005**, *48*, 7315.
- [65] Capiro, V.; Guyen, B.; Opoku-Boahen, Y.; Mann, Y.; Gowan, S. M.; Kelland, L. M.; Read, M. A.; Neidle, S. *Bioorg. Med. Chem. Lett*., **2000**, *10*, 2063.
- [66] Guyen, B.; Schultes, C. M.; Haze, P.; Mann, J.; Neidle, S. *Org. Biomol. Chem.,* **2004**, *2*, 981.

Copyright of Mini Reviews in Medicinal Chemistry is the property of Bentham Science Publishers Ltd. and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.